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1. Product description

Components and specifications

Chemically defined
Free of animal-derived components
Free of proteins

without L-glutamine
without HT (hypoxanthine/thymidine)
without Growth Factor

Storage

Store protected from light at 2–8 °C.
Do not freeze.

Intended use

Intended for *in vitro* research and manufacturing processes **only**. Do not use for injection or infusion!

2. Background information and applications

CH0lean Medium is a complete chemically-defined, animal-component-free, protein-free, and ready-to-use medium. CH0lean is a lean medium formulation designed for batch and fed-batch processes with different CHO cell lines. CH0lean is developed with a minimal set of well-balanced components to achieve a lean, robust and flexible medium basis. In addition, it is capable of competing with state-of-the-art media for highest cell growth and product titer. The formulation is suitable for a broad range of different CHO cell lines, including K1 and DG44. Furthermore, CH0lean is optimized for easy adaptation from other media and is a robust platform to build high-performance fed-batch processes on. The medium supports cell growth and production of recombinant proteins and antibodies in

suspension culture. It can be used in research or in manufacturing applications.

3. Protocols

3.1 Preparations

All procedures should be carried out using sterile techniques in a biosafety cabinet.

CH0lean is formulated without L-glutamine. For applications requiring this amino acid, supplement with 6-8 mM L-glutamine prior to use. Supplementation of L-glutamine directly to the culture is recommended.

3.2 Culture conditions

Cultures should be maintained at 37 °C. For cultivation in an incubator, a 5% CO₂ atmosphere is necessary.

Parameter	Value [-]
Orbital shaker deflection	5 cm
Shaking speed	130-185 rpm
Temperature	37°C
CO ₂	5%

Table 1: Recommended culture conditions for use of Xcell media and feed products.

Using the set-up listed in table 1, the working volume of different polycarbonate Erlenmeyer shake flask sizes was determined (table 2). For cell lines with strong aggregation, the addition of anti-clumping agent or Poloxamer 188 (1-3 g/L) could be tested or baffled shake flasks may be used. For this setup, a reduction of the shaking speed might be necessary.

Size of shaker [mL]	Shape [-]	Working volume [mL]
125	plain, vent cap	25 - 50
250	plain, vent cap	80 - 150
500	plain, vent cap	200 - 300
1000	plain, vent cap	400 - 600

Table 2: Recommended culture working volumes for use of Xcell media and feed products in various shake flask sizes.

3.3 Instructions for use

3.3.1 Thawing of cells

- 1) Quickly thaw a vial of frozen cells in a 37 °C water bath.
- 2) Transfer the cells aseptically to a centrifugation tube containing 10 mL of CH0lean Medium or PBS buffer.
- 3) Centrifuge cell suspension at 115×g for 5 minutes.
- 4) Aspirate supernatant completely and discard.

- 5) Resuspend the cells in a sufficient amount of CHOlean Medium per vial (e.g. 20 mL Medium for 1 vial of 1×10^7 cells).
- 6) Adjust viable cell density to $5\text{-}10 \times 10^5$ cells/mL by medium addition and transfer cell suspension into your cultivation system (e.g. 125 mL polycarbonate Erlenmeyer flask, or 50 mL tube spin bioreactor).
- 7) Count the cells after 24-48 hours for assessment of cell density and viability.
- 8) Adjust cell density to $3\text{-}6 \times 10^5$ cells/mL. *
- 9) Proceed with routine cultivation.

* Depending on the cell line, the target inoculation cell density can be lower.

3.3.2 Routine cultivation and cell expansion

- 1) Pre-equilibrate a sufficient amount of medium in a polycarbonate Erlenmeyer shake flask (Parameters listed in tables 1 and 2) for 1 hour. **
- 2) Determine viable cell density in the pre-culture.
- 3) Depending on the inoculation volume, remove medium from the shake flask to reach the target working volume after inoculation. Final working volume of given shaker size is listed in table 2.
- 4) Seed cells at a target inoculation cell density of 3×10^5 cells/mL (operational range $2\text{-}5 \times 10^5$ cells/mL).
- 5) Incubate the culture according to the conditions listed in table 1.
- 6) Routinely passage the culture when viable cell densities between $15\text{-}40 \times 10^5$ cells/mL are reached. Typical duration time for the culture is 3-4 days.
- 7) If cell density is too low or cells do not grow in adaption phase, centrifuge the culture and exchange the medium without dilution after 4 days.

** Depending on cell line, the pre-equilibration of medium might be not necessary. For some cell lines the use of 2-8°C cold culture medium directly from refrigerator was found to be beneficial. This procedure eliminates handling variations of the medium in the pre-equilibration phase of the medium.

3.3.3 Stepwise adaptation from serum-containing cultures

- 1) Expand the culture in serum-containing standard medium.
- 2) Centrifuge a sufficient number of cells for inoculation of suspension culture with $4\text{-}6 \times 10^5$ cells/mL at $115 \times g$ for 5 minutes.
- 3) Resuspend cells in Xell medium (if necessary, include 6-8 mM L-glutamine) and 2 % fetal bovine serum (FBS).
- 4) Passage cells or change medium by centrifugation every two to four days depending on cell density.
- 5) Reduce serum concentration to 0.5 % after at least three passages.
- 6) Passage cells or change media by centrifugation every two to four days depending on cell density.
- 7) Reduce serum concentration to 0 % after two to four passages.
- 8) Continue cultures until viabilities stabilize at > 90 %.
- 9) Adapted cells should be inoculated at $2\text{-}5 \times 10^5$ cells/mL in Xell medium for optimal performance. Cultures should be diluted every three or four days. Due to aggregation of CHO cells,

cultures should be stirred or shaken, using spinner bottles, shaker flasks or similar cultivation systems.

3.3.4 Bioreactor cultivation

For best performance the inoculation density in bioreactor should be in the range of $3\text{-}5 \times 10^5$ cells/mL in Xell medium. Suggested starting parameters for bioreactor cultivations of CHO cells using Xell medium are pH 7.1-7.6, 30-40% DO, and a temperature of 37 °C. The medium already contains Poloxamer188, further supplementation is not necessary. For fed-batch processes, CHOlean medium can be combined with e.g. Basic Feed or TCX7D Feed.

3.3.5 Freezing of cells

Cells can be frozen in CHOlean Medium without the use of serum.

- 1) Choose a well-growing culture with viabilities above 90 %.
- 2) Prepare a freezing medium consisting of 90 % CHOlean Medium and 10 % dimethyl sulfoxide (DMSO; cell culture grade).
- 3) Cool down the freezing medium to 2-8 °C.
- 4) Centrifuge the cells at $115 \times g$ for 5 minutes.
- 5) Aspirate supernatant completely.
- 6) Resuspend the cells in freezing medium at 1×10^7 cells/vial.
- 7) Rapidly transfer 1.5 mL of this suspension to sterile cryovials.
- 8) Place the vials in a pre-cooled (2-8 °C) freezing module and store the modules including the vials for 24 hours at -80 °C.
- 9) Transfer the cryovials to a -140 °C to -196 °C system for long time storage.

4. Ordering information

Product	Application	Order No.
TC-42	base medium and protein production for various CHO cells; esp. CHO DHFR cell lines	509 (rInsulin) 510 (IGF) 511 (w/o GF)
TCX6D	base medium and protein production for various CHO cells	1070
TCX10D	base medium and protein production for various CHO cells; esp. for CHO GS cell lines	1100
CHOlean	base medium and protein production for various CHO cells	1140
CHO TF	transfection medium, suitable for growth, transfection/infection and production for various CHO cells; esp. for transfection	886
Basic Feed	feed supplement for CHO, HEK, hybridoma, <i>et al.</i>	1092
TCX7D Feed	Feed supplement for various CHO cells	1080

Table 3: CHO products by Xell

Place orders: order@xell.de

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